

RESEARCH ARTICLE

Molecular and Morphological Characterization of Three New Fungal Species of the Class Sordariomycetes from Korea

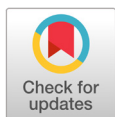
Kallol Das¹, Jae-Ho Ban^{1,2}, So-Young Choi¹, Seung-Yeol Lee^{1,3,*}, and Hee-Young Jung^{1,3}¹College of Agriculture and Life Sciences, Kyungpook National University, Daegu 41566, Korea²Plant Quarantine Technology Center, Animal and Plant Quarantine Agency, Gimcheon 39660, Korea³Institute of Plant Medicine, Kyungpook National University, Daegu 41566, Korea

*Corresponding author: leesy1123@knu.ac.kr

ABSTRACT

Three fungal strains belonging to the class Sordariomycetes were isolated from soil collected on Jeju Island and Gyeongsangbuk-do, Korea. They were identified as *Diaporthe endophytica* (KNU-JJ-1809), *Faurelina indica* (KNU-JJ-1830), and *Trichoderma ivoriense* (KNU-4-KH1). KNU-JJ-1809 produced beta conidia that were straight, curved, hyaline, smooth-walled, with a diameter of $16.5\text{--}25.0 \times 0.6\text{--}1.7\ \mu\text{m}$. The conidia of strain KNU-JJ-1830 were hyaline to light green, thin, clavate, round, truncate base, had guttules at both ends, with a diameter of $2.5\text{--}5.2 \times 1.7\text{--}3.8\ \mu\text{m}$. The conidia of strain KNU-4-KH1 were oblong or ellipsoidal, smooth-walled, greenish, with a diameter of $2.2\text{--}4.4 \times 2.2\text{--}3.6\ \mu\text{m}$. Internal transcribed spacer regions, partial large subunit, translation elongation factor 1- α , β -tubulin, and calmodulin genes were used to confirm the strains, and their cultural and morphological characteristics. To our knowledge, this is the first report on *D. endophytica*, *F. indica*, and *T. ivoriense* in Korea.

Keywords: *Diaporthe endophytica*, *Faurelina indica*, Sordariomycetes, *Trichoderma ivoriense*



OPEN ACCESS

pISSN : 0253-651X
eISSN : 2383-5249

Kor. J. Mycol. 2023 March, 51(1): 25-38
<https://doi.org/10.4489/KJM.20230003>

Received: March 09, 2023

Revised: March 15, 2023

Accepted: March 20, 2023

© 2023 THE KOREAN SOCIETY OF MYCOLOGY.



This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

INTRODUCTION

The class Sordariomycetes includes taxa with perithecial ascomata, paraphysate hamathecium, paraphysate ostioles, and unitunicate or pseudoprotunicate asci [1]. Sordariomycetes is the second largest class of Ascomycota and contains 1,331 genera and 105 families, including 32 orders and 6 subclasses [2]. The genus *Diaporthe*, belonging to the class Sordariomycetes, was identified by Nitschke, using the type species *D. eres* from *Ulmus* sp. collected in Germany. It belongs to the family Diaporthaceae and order Diaporthales [2,3]. Numerous species names in the genus *Diaporthe* have emerged over the years because of their introduction, which is mostly based on host relationships. A single species can be found in one or more hosts, and it is now understood that the species are not host-specific [4]. The key factor used to classify different *Diaporthe* species is their molecular phylogeny, particularly those based on the internal transcribed

spacer (ITS) of nuclear ribosomal DNA. However, some authors have also used the translation elongation factor 1- α (*TEF1 α*) gene [5]. The classification of the genus relies heavily on molecular phylogenies, particularly those generated from sequences of the ITS, *TEF1 α* , β -tubulin (*TUB2*), and calmodulin (*CAL*) loci [6]. Another member of the class Sordariomycetes, belonging to the genus *Faurelina* was first described as a new genus of cleistothecial ascomycetes [7]. This genus has a problematic taxonomic history. Initially, *Faurelina* was tentatively placed in Chadeaudiellaceae; however, this position was questionable. The type species *Faurelina* belongs to the class ascolocular ascomycetes and should be placed in the class formerly known as Loculoascomycetes [8]. *Faurelina* was then transferred to Microascaceae after two years, mainly based on the presence of dextrinoid ascospores without germ pores [9]. The placement of *Faurelina* in Microascales partly resulted from the study by [10], who sequenced the nucLSU, nucSSU, and *RPB2* gene regions from a single strain labeled *F. indica* (CBS 126.78), giving results identical to those of the type species *Ceratocystis fimbriata*.

In addition, the well-known genus belonging to the class Sordariomycetes, called *Trichoderma*, is distinguished by perithecia submerged in fleshy stromata of various colors, cylindrical asci with green or hyaline ascospores that disarticulate at the septum, conidiophores that create various branch patterns, and green or hyaline conidia [11]. The taxonomic classification of *Trichoderma* spp. using traditional methods is challenging. Recently, the study of genus taxonomy has advanced based on combined assessments of morphology and DNA sequence data, demonstrating that the color of ascospores can reveal important evolutionary aspects [12]. Although *Trichoderma* is a well-studied genus, there are several species whose evolutionary positions are not fully understood [13].

The identification and classification of fungal species isolated from Korean soil was the main goal of this extensive investigation. Fungal species, together with their morphological and cultural traits, were identified using molecular phylogenetic analyses. In this study, three fungal species from Korea are described and illustrated as new records.

MATERIAL AND METHODS

Soil sampling and fungal isolation

In 2018 and 2019, soil samples were collected from Jeju Island (33°25'22.9"N, 126°33'01.6"E) and Gyeongsangbuk-do (35°51'39.7"N, 129°16'09.5"E), South Korea. Using a sterile pre-autoclaved spatula, soils from depths of 10-30 cm were randomly sampled, transferred to sterile zipper plastic bags, and then transported to the laboratory. The soil samples (1 g) were suspended in 10 mL of sterile distilled water, vortexed gently, and serially diluted, and a volume of 100 μ L was spread on potato dextrose agar (PDA; Difco, Detroit, MI, USA) plates and incubated for 2-3 days at 25°C. The germinating single colonies were then transferred to new PDA plates at 25°C. A pure culture of the strain was selected for further molecular analysis based on specific characteristics. For further study, the fungal strains were maintained in 20%

glycerol at -80°C. Stock cultures of three strains, namely KNU-JJ-1830 (NIBRFGC000502239), KNU-JJ-1809 (NIBRFGC000502235), and KNU-4-KH1 (NIBRFGC000505959), were deposited at the National Institute of Biological Resources (NIBR) as metabolically inactive cultures.

Cultural and morphological characterization

Cultural and morphological characteristics were evaluated using different media, depending on the genus and species. KNU-JJ-1809 and KNU-JJ-1830 were cultured on PDA, oatmeal agar (OA; Difco, Detroit, MI, USA), and malt extract agar (MEA; Difco, Detroit, MI, USA) and incubated at 25°C for 2 and 3 weeks, respectively [6,14]. KNU-4-KH1 was transferred on PDA, MEA, cornmeal dextrose agar (CMAD: cornmeal, 40.0 g/L; dextrose, 20.0 g/L; agar, 15.0 g/L; distilled water, 1,000 mL), and synthetic nutrient agar (SNA) followed by 2 weeks at 25°C [15,16]. Next, cultural features such as colony color, fungal development, and texture, were documented. Mycological characteristics of the fungal strains were observed under a light microscope (BX-50, Olympus, Tokyo, Japan).

Genomic DNA extraction, PCR amplification, and sequencing

Total genomic DNA was extracted using the HiGene Genomic DNA Prep Kit (BIOFACT, Daejeon, Korea) according to the manufacturer's instructions. The PCR amplification process was conducted using a fragment of the ITS regions as a target using primers ITS1F/ITS4 [17,18]; 28S ribosomal DNA of large subunit using LROR/LR5 [19]; *TEF1* α (EF1-728F/TEF1rev or EF1-986R) [20,21]; *CAL* (CAL-228F/CAL-737R) [22]; and *TUB2* (T1/Bt-2b) [22,23]. The products were run on an ethidium bromide-stained 1% agarose gel to confirm their yields, purified using EXOSAP-IT (Thermo Fisher Scientific, Waltham, MA, USA), and sequenced by Solgent (Daejeon, Korea).

Molecular phylogenetic analysis

For phylogenetic analyses, the related sequences were retrieved from the National Center for Biotechnology Information (Table 1). The MEGA7.0 program was used to align each gene and combine the sequences to show where the strains fell on the phylogenetic tree. Ambiguous regions were deleted from the alignments, and the evolutionary distance matrices for the neighbor-joining (NJ) algorithm were calculated using Kimura's two-parameter model [24]. The NJ method was inferred from the tree topology using MEGA7 with bootstrap values based on 1,000 replications [25].

Table 1. Strains and GenBank accession numbers used for phylogenetic analysis in this study.

Species	Strain numbers	GenBank accession numbers				
		ITS	<i>TEF1α</i>	<i>CAL</i>	<i>TUB2</i>	LSU
<i>Diaporthe batatas</i>	CBS 122.21 ^T	NR152456	KC343766	KC343282	KC344008	-
<i>Diaporthe citri</i>	CBS 199.39	KC343051	KC343777	KC343293	KC344019	-
<i>Diaporthe convolvuli</i>	CBS 124654	KC343054	KC343780	KC343296	KC344022	-
<i>Diaporthe corylina</i>	CBS121124	KC343004	KC343730	KC343246	KC343972	-
<i>Diaporthe endophytica</i>	LGMF948	KC343072	KC343798	KC343314	KC344040	-
<i>Diaporthe endophytica</i>	LGMF935	KC343070	KC343796	KC343312	KC344038	-
<i>Diaporthe endophytica</i>	LGMF919	KC343067	KC343793	KC343309	KC344035	-
<i>Diaporthe endophytica</i>	CBS 133811 ^T	NR111847	KC343791	KC343307	KC344033	-
<i>Diaporthe endophytica</i>	KNU-JJ-1809	LC567429	LC567434	LC567433	LC567435	-
<i>Diaporthe melonis</i>	CBS 435.87	KC343141	KC343867	KC343383	KC344109	-
<i>Diaporthe phaseolorum</i>	CBS 116019	KC343175	KC343901	KC343417	KC344143	-
<i>Diaporthe sojae</i>	CBS 180.55	KC343200	KC343926	KC343442	KC344168	-
<i>Chlamydotubeufia huaikangplaensis</i>	MFLUCC10-0926	-	-	-	-	JN865198
<i>Faurelina indica</i>	CBS 126.78	-	-	-	-	GU180653
<i>Faurelina indica</i>	CBS 301.78	-	-	-	-	GU180654
<i>Faurelina indica</i>	KNU-JJ-1830	LC567431	-	-	-	LC567432
<i>Hysterographium fraxini</i>	CBS 109.43	-	-	-	-	FJ161171
<i>Ostreichnion curtisii</i>	CBS 198.34	-	-	-	-	FJ161176
<i>Rhexothecium globosum</i>	CBS 955.73 ^T	-	-	-	-	HG004544
<i>Trichoderma barbatum</i>	CBS 125733 ^T	HQ342417	HQ342223	HQ342352	-	-
<i>Trichoderma caesareum</i>	CBS 124369 ^T	HQ342410	HQ342216	HQ342345	-	-
<i>Trichoderma floccosum</i>	CBS 124372 ^T	HQ342412	HQ342218	HQ342347	-	-
<i>Trichoderma ivoriense</i>	CBS 125734 ^T	HQ342411	HQ342217	HQ342346	-	-
<i>Trichoderma ivoriense</i>	KNU-4-KH1	LC567430	LC567437	LC567436	-	-
<i>Trichoderma lanuginosum</i>	CBS 125718 ^T	HQ342415	HQ342221	HQ342350	-	-
<i>Trichoderma medusae</i>	CBS 125719 ^T	HQ342408	HQ342214	HQ342343	-	-
<i>Trichoderma rossicum</i>	DAOM 230011 ^T	HQ342419	AY937441	HQ342354	-	-
<i>Trichoderma vermipilum</i>	CBS 127103 ^T	HQ342413	HQ342219	HQ342348	-	-
<i>Trichoderma virens</i>	DIS 162	FJ442669	FJ463367	FJ442403	-	-

The strains identified in this study are indicated in bold. ^T, indicates type strain.

ITS: internal transcribed spacer regions; *TEF1α*: translation elongation factor 1-alpha; *CAL*: calmodulin; *TUB2*: β-tubulin; LSU: 28S ribosomal DNA.

RESULTS AND DISCUSSION

Taxonomy

Diaporthe endophytica R. R. Gomes, C. Glienke & Crous, Persoonia 31:20 (2013) (Fig. 1).

Ecology and Distribution: Most members of this genus have been recorded from different hosts, including endophytic, saprobic, and plant-pathogenic fungi such as the leaves of *Acacia retinodes* (Australia), branches of *Betula alleghaniensis* (Canada), *Pyrus communis* (South Africa), *Helianthus annuus* (Italy), twigs of *Ulmus* sp. (Germany), stems and twigs of *Toxicodendron vernicifluum* (Japan), and soil (Korea). The strain KNU-JJ-1809 was collected from Korean soil.

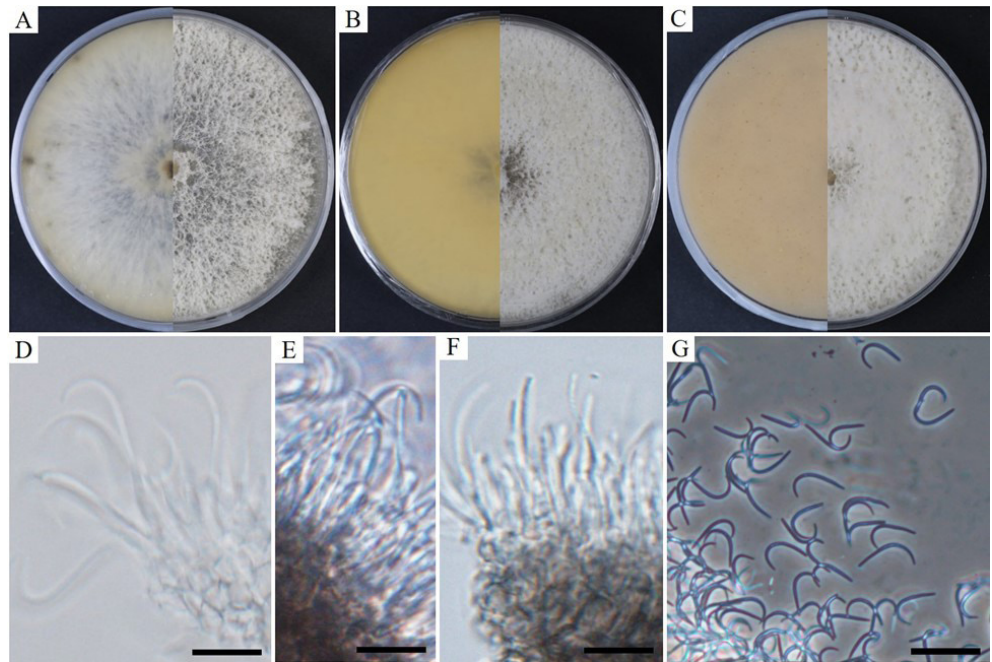


Fig. 1. Cultural and morphological characteristics of *Diaporthe endophytica* KNU-JJ-1809. Colony on potato dextrose agar (A); malt extract agar (B); and oatmeal agar (C). Conidiogenous cells (D-F); beta conidia (G). Scale bars: D-G=10 µm.

Cultural and morphological characteristics: The strains were cultured on PDA, MEA, and OA media to study their cultural and morphological characteristics (Table 2). The strain produced sparse aerial mycelium, buff, white to isabelline, reverse smoke-grey, covering the petri dish after 2 weeks on PDA at 25°C (Fig. 1A). The colonies were sparse, buff, white to smoke-grey, reverse grey to olivaceous-grey, and covered the petri dish after 2 weeks on MEA at 25°C (Fig. 1B) and OA gray to smoke-gray (Fig. 1C). The conidiophores were hyaline, branched, densely aggregated, cylindrical, straight to curved, and had 1–3-septate. Conidiogenous cells were hyaline, subcylindrical and filiform, straight, slightly tapering toward the apex, and with a diameter of (n=10) $14.7\text{--}24.4 \times 1.1\text{--}2.3$ µm (average, 18.7×1.5 µm) (Figs. 1D-F)). No alpha-conidia were observed. Beta conidia were commonly found, straight, curved, hyaline, smooth-walled, and with a diameter (n=30) of $16.5\text{--}25.0 \times 0.6\text{--}1.7$ µm (average= 20.1×1.2 µm) (Fig. 1G). Based on the cultural and morphological characteristics, KNU-JJ-1809 was similar to the previously identified *D. endophytica* (Table 2).

Table 2. Morphological characteristics of the strain KNU-JJ-1809 with reference to *Diaporthe endophytica*.

Characteristics		KNU-JJ-1809 ^a	<i>Diaporthe endophytica</i> CBS 133811 ^b
Colony	Shape, color, and size (diam.)	PDA: sparse aerial mycelium, buff, white to isabelline; reverse smoke-grey. MEA: sparse, buff, white to smoke-grey; reverse grey to olivaceous-grey. OA: grey to smoke-grey.	PDA: sparse aerial mycelium, buff, honey to isabelline; reverse smoke-grey. MEA: buff with umber patches; reverse dark mouse-grey, with patches of isabelline. OA: smoke-grey to olivaceous-grey.
Conidiogenous cells	Shape and size (diam.)	Hyaline, subcylindrical and filiform, straight, slightly tapering towards the apex, $14.7\text{--}24.4 \times 1.1\text{--}2.3$ µm.	N/A
Alpha conidia		N/A	N/A
Beta conidia	Shape	Hyaline, smooth, curved.	N/A
	Size (diam.)	$16.5\text{--}25.0 \times 0.6\text{--}1.7$ µm (n=30)	N/A

N/A: not available in the literature; PDA; Potato dextrose agar; MEA: Malt extract agar; OA: Oatmeal agar.

^a Fungal strain used in this study, ^b Source of description [6].

Molecular phylogeny of KNU-JJ-1809

Sequence analysis revealed that 485 bp of the ITS region, 440 bp of *CAL*, 305 bp of *TEF1α*, and 445 bp of *TUB2* were obtained. The BLAST search results of the ITS sequences revealed that KNU-JJ-1809 exhibited 100% similarity with different strains of *D. endophytica* (CBS 133811^T, NOU760, NTOU 4884, NTOU 4380, NTOU 4920, PHA-1, and ZJUD73). *CAL* showed maximum 98.2-98.9% similarity with the *D. endophytica* strains (LGMF937, CBS 133811^T, and LGMF935). *TEF1α* showed maximum 99.3-99.7% similarity with the *D. endophytica* strains (SoyYS1732, ZJUD94, and ZJUD73), and the *TUB2* displayed maximum 99.2% similarity with the *D. endophytica* strain LGMF935. Phylogenetic analysis was performed based on a combination of ITS, *CAL*, *TEF1α*, and *TUB2* sequences using the NJ phylogenetic tree. Analysis of the combined sequences in the phylogenetic tree revealed that KNU-JJ-1809 clustered with a previously identified strain of *D. endophytica* (Fig. 2). Therefore, KNU-JJ-1809 was identified as *D. endophytica*.

According to previous research, *Diaporthe* species are common in temperate and tropical regions as endophytes associated with several hosts [26]. *D. betulae* and *D. betulicola* have been identified on the genus *Betula* in China, and often appear to be the source of birch dieback with typical symptoms. Although multiple plant pathogenic *Diaporthe* species have been identified in China, little is known about economically and ecologically valuable species [27]. In addition, *D. phaseolorum* is an endophyte found in mangrove forests [28]. The closely related taxa *D. endophytica*, *D. schini*, and *D. terebinthifolii* were originally described as endophytes from *Schinus* spp. in Brazil [6]. Additionally, *Diaporthe* spp. are known to be infectious, particularly in people with weakened immune systems; however, they are rare diseases of major medical importance [29]. *D. endophytica* was found to be an endophytic from the medicinal plants *S. terebinthifolius* and *Maytenus ilicifolia* in Brazil [6], however, *D. endophytica* can control the pathogen *Phyllosticta citricarpa* that is responsible for citrus black spot in citrus plants [30]. However, *Diaporthe* species associated with plant and forest tree diseases have rarely been studied in Korea. Therefore, specimens must be collected from a wide variety of habitats and hosts to fully understand the taxonomy of *Diaporthe* species. KNU-JJ-1809 was isolated from soil and identified as *D. endophytica* which has not been previously reported in Korea.

Taxonomy

Faurelina indica Arx, Mukerji & N. Singh, Sydowia 34:39 (1981) (Fig. 3).

Ecology and distribution: Very few members of this genus have been reported to date. Some species were identified in the fruit of herbivore dung collected from the Caatinga biome in the semi-arid region of Brazil (*F. fimigena* and *F. hispanica*). The type species *F. indica* was isolated from the dung of cows and goats (India). Some members of this genus have been identified in Japan and France. The present studied strain KNU-JJ-1830 was collected from soil in Korea.

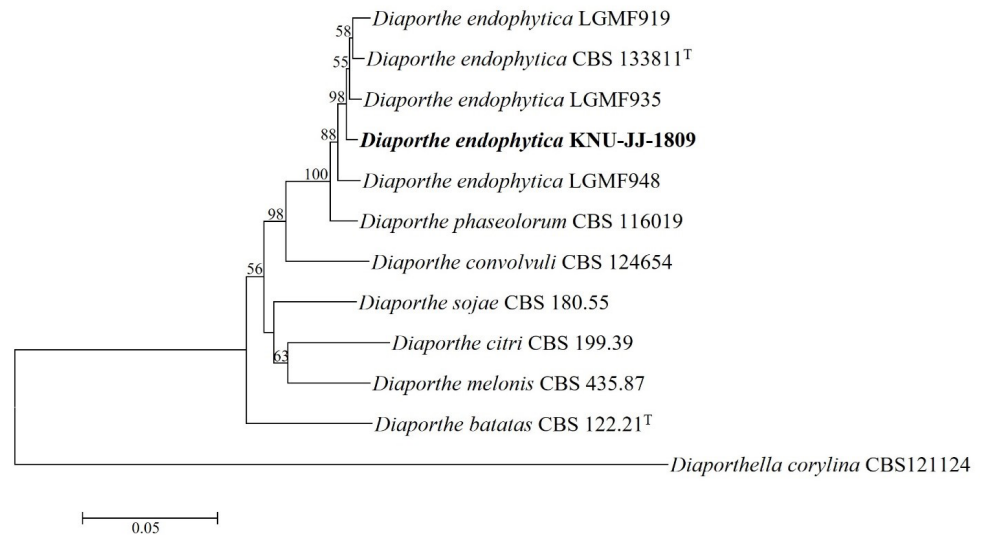


Fig. 2. Neighbor-joining phylogenetic tree of the strain KNU-JJ-1809 based on the combined sequences (ITS+*TUB2*+*CAL*+*TEF1α*), showing the relationships between *Diaporthe endophytica* with the closest species of *Diaporthe*. *Diaporthella corylina* CBS 121124 was used as an outgroup. Numbers above the branches represent the bootstrap values obtained for 1,000 replicates (values smaller than 50% are not shown). The isolated strain of this study is indicated in bold. Bar, 0.05 substitutions per nucleotide position. ITS, internal transcribed spacer of nuclear ribosomal DNA; *TUB2*, β-tubulin gene; *CAL*, calmodulin gene; *TEF1α*, translation elongation factor 1-alpha gene.

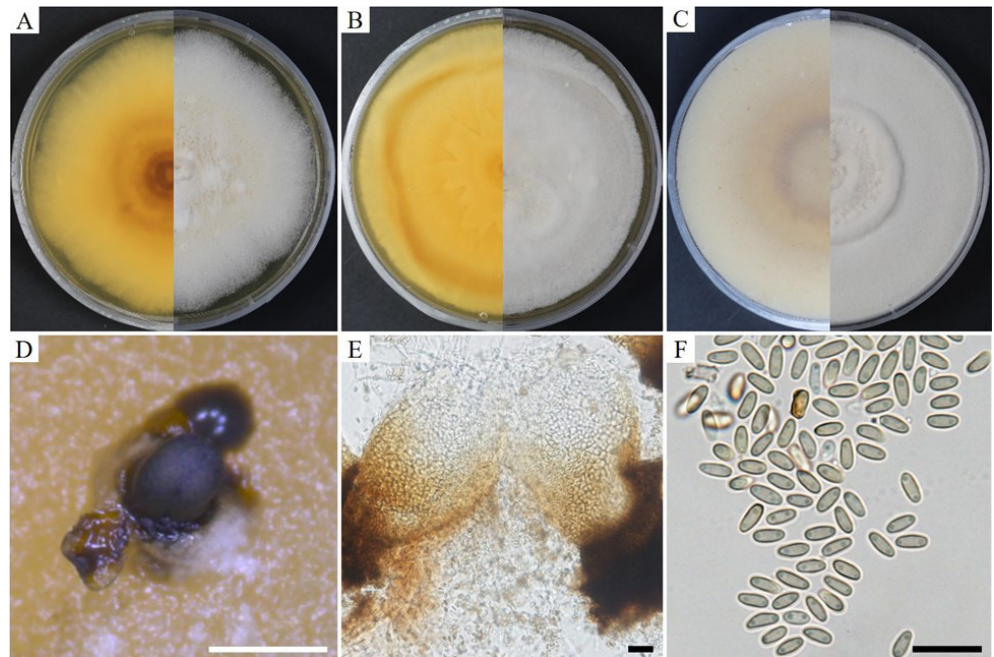


Fig. 3. Cultural and morphological characteristics of *Faurelina indica* KNU-JJ-1830. Colony on potato dextrose agar (A); malt extract agar (B); and oatmeal agar (C); Conidiomata forming on OA (D); pycnidial wall releasing mature conidia (E); and conidia (F). Scale bars: D=500 μm; E=50 μm; F=10 μm.

Cultural and morphological characteristics: To study the cultural and morphological characteristics, the strain was cultured on PDA, MEA, and OA media (Table 3). The colonies were white to pale yellow, reaching 80-85 mm in diameter on PDA after 21 days at 25°C, and reverse yellowish to dark yellow toward the center (Fig. 3A). On MEA, the colonies were white to pale yellow, reverse yellow, yellow patches toward the edge, and irregular patches near the center, covering 85-90 mm in diameter after 21 days at 25°C (Fig. 3B). On OA, the colonies were white, smooth, with tufts of irregular aerial mycelia toward the center, reverse light brown to brown toward the center, and covered the Petri dishes after 21 days at 25°C (Fig. 3C). Conidiomata were immersed at the base, dark black, hemispherical or pustulate, rounded above, smooth, and 200-350 µm in size (Fig. 3D). The vegetative hyphae were hyaline or light brown, thick-walled, branched, and septate. The pycnidial walls released mature yellowish conidia (Fig. 3E). Conidia were hyaline to light green, thin, clavate, round, truncate base, with guttules in both ends, and with a diameter (n=100) of $2.5\text{--}5.2 \times 1.7\text{--}3.8$ µm (average, 4.5×2.0 µm) (Fig. 3F). Based on previous studies, KNU-JJ-1830 was found to be similar to *F. indica* based on its cultural and morphological characteristics (Table 3).

Table 3. Morphological characteristics of the strain KNU-JJ-1830 with reference to *Faurelina indica*.

Characteristics		KNU-JJ-1830 ^a	<i>Faurelina indica</i> CBS 126.78 ^b
Colony	Shape and size (diam.)	OA: white, smooth, tufts of irregular; reverse light brown to brown covering petri dishes after 21 days at 25°C.	OA: dark brown, with irregular tufts of aerial mycelium, with daily growth rate 1-1.5 mm at 20°C.
	Hyphae	Vegetative hyphae were hyaline or light brown, thick-walled, branched, septate.	Vegetative hyphae thick-walled, hyaline, or light brown.
Conidia (Arthroconidia)	Shape	Hyaline to light green, thin, clavate, round, truncate base, guttules in both ends	Arthroconidia either directly or on shorter or longer stalks, clavate, with a truncate base, hyaline.
	Size (diam.)	$2.5\text{--}5.2 \times 1.7\text{--}3.8$ µm.	$3.0\text{--}5.0 \times 1.5\text{--}2.5$ µm.

N/A: not available in the literature; OA: Oatmeal agar.

^a Fungal strain used in this study, ^b Source of description [14].

Molecular phylogeny of KNU-JJ-1830

Nucleotide sequences of the ITS (547 bp) and 28S ribosomal DNA (883 bp) regions were obtained after sequence analysis. The 28S rDNA showed 98.98% and 99.21% identity with *F. indica* strains CBS 126.78 and CBS 301.78, respectively. The 28S ribosomal DNA dataset obtained using the NJ phylogenetic algorithm showed that KNU-JJ-1830 and the reference strains of *F. indica* (CBS 126.78 and CBS 301.78) produced clusters with a high bootstrap value (100%) (Fig. 4). Thus, KNU-JJ-1830 was identified as *F. indica*.

The type species of *F. indica* and *F. elongata* (previously known as *Leuconeurospora elongata*) were previously identified in goat and cow dung collected in Nainital, Delhi, India [12]. Although one of these species was *L. elongata*, it was later transferred to the genus *Faurelina* [10]. *F. elongata* was obtained from Kagoshima (southwestern Kyushu, Japan). Moreover, the fruiting of two species, *F. fimigena* and *F. hispanica*, was recorded in herbivore dung collected from the Caatinga biome in the semi-arid region

of Brazil [31]. However, a lack of knowledge about fungal diversity, particularly coprophilous fungi, is an important factor that complicates conservation strategies. A focused study of herbivore dung mycobiota can improve our knowledge of the biogeography and diversity of numerous unreported species. Few species have been identified in the genus *Faurelina* and have been recorded worldwide. This is the first recorded genus and the first record of *F. indica* in Korea.

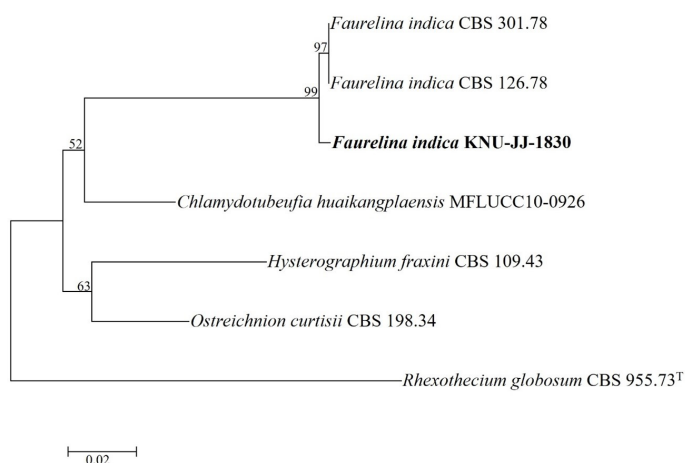


Fig. 4. Neighbor-joining phylogenetic tree of the strain KNU-JJ-1830 based on sequences of 28S ribosomal DNA to reveal the phylogenetic position of *Faurelina indica*. *Rhexothecium globosum* CBS 955.73^T was used as an outgroup. Numbers above the branches represent the bootstrap values obtained for 1,000 replicates. The isolated strain of this study is indicated in bold. Bar, 0.02 substitutions per nucleotide position.

Taxonomy

Trichoderma ivoriense Samuels, Mycological Progress 11:237 (2011) (Fig. 5).

Ecology and distribution: Members of this genus have been observed on several hosts, such as mushroom compost (Canada, Ireland, England, USA), soil in kiwifruit orchards (New Zealand), forest soil (USA), *Quercus* infected with *Lentinus edodes* (Japan), botanical garden soil (England), maize field soil (China, Colombia), *Pleurotus* spawn (Netherlands), soil (Japan, Cameroon), water-damaged buildings (Denmark), and vegetable greenhouse soil (China). The present studied strain KNU-4-KH1 was isolated from soil in Korea.

Cultural and morphological characteristics: KNU-4-KH1 was cultured at 25°C for 72 h in PDA, CMAD, MEA, or SNA media (Table 4). Consequently, mycelial growth was 74.4-77.7, 74.0-90.0, 67.5-71.5, and 82.0-90.0 mm, respectively. The mycelia initially appeared white, turned pale green, and later became green on PDA (Fig. 5A). The hyphae were sparse and hyaline to greenish with two concentric circles, which became thicker. Several green and white pustules were observed in the medium. CMAD did not change color to white hyphae, even after one month (Fig. 5B). On MEA, the mycelia turned from white to very pale green over time, gradually turned yellow, and had two concentric circles, similar to that on PDA (Fig. 5C). On SNA medium, the hyphae grew close to the medium surface, and green pustules

were observed around the incised area (Fig. 5D). Conidiophores in the pustules had long, sterile hairs that were light brown to blackish in color and short, fertile branches growing from their bases (Figs. 5E and F). Fertile branches growing from the base of the hair normally had a single basal cell and produced terminal phialides, whereas branches farther from the tip typically did not have a single basal cell and did not produce terminal phialides (Figs. 5G and H). Numerous conidia were observed on the PDA. Conidia were oblong or ellipsoidal, smooth-walled, greenish, and with a diameter (n=50) of $2.0\text{--}4.4 \times 2.2\text{--}3.6\text{ }\mu\text{m}$ (Fig. 5I). Strain KNU-4-KH1 was confirmed to be a previously identified *T. ivoriense* strain based on its cultural and morphological characteristics (Table 4).

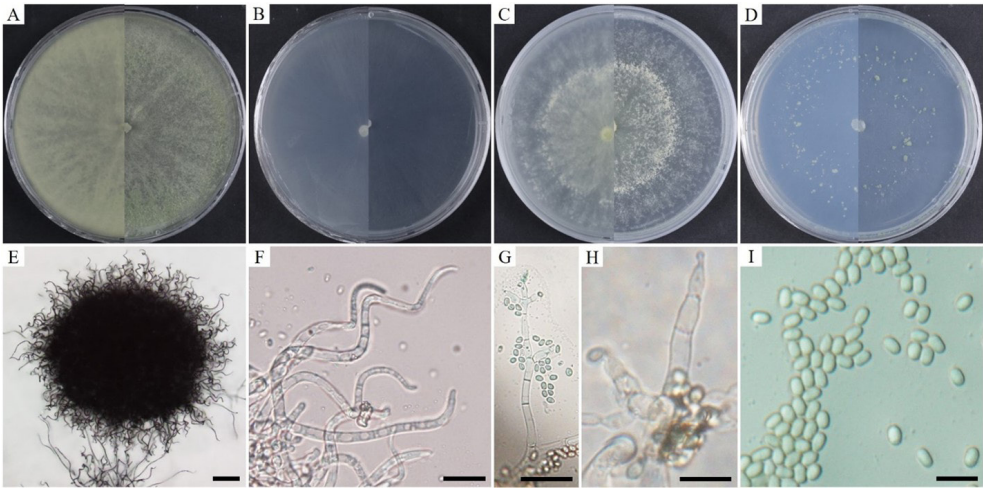


Fig. 5. Cultural and morphological characteristics of *Trichoderma ivoriense* KNU-4-KH1. Colony on potato dextrose agar (A); corn meal agar (B); malt extract agar (C); and synthetic nutrient-poor agar (D); Hairs on conidial pustules (E, F); fertile branch with phialides (G, H); and conidia (I). Scale bars: E=100 μm ; F-I=10 μm .

Table 4. Morphological characteristics of the strain KNU-4-KH1 with reference to *Trichoderma ivoriense*.

Characteristics		KNU-4-KH1 ^a	<i>Trichoderma ivoriense</i> BJ598-1 ^b
Colony	Color	PDA: white mycelium, turn to green; MEA: white turn to green and change to yellow; SNA: white mycelium; CMAD: white mycelium.	PDA: white at first and slowly turning bright glaucous; MEA: conidiation compact green.
	Size (diam.)	PDA: 74-90 mm; CMAD: 74-77 mm; MEA: 67-71 mm; SNA: 76-90 mm after 72h.	PDA: 66-72 mm; CMAD: 70-76 mm; MEA: 66-72 mm; SNA: 68-74 mm after 72 h.
	Shape	PDA: aerial mycelium sparse, concentric ring; CMAD: aerial mycelium sparse; MEA: aerial mycelium sparse, concentric ring.	PDA: aerial mycelium sparse, concentric ring; CMAD: aerial mycelium sparse; MEA: aerial mycelium sparse, concentric ring.
Phialide	Shape	PDA: slender	PDA: slender, subulate.
Conidia	Shape	PDA: ellipsoid, smooth-walled	PDA: oblong or ellipsoid, smooth-walled, subhyaline.
	Size (diam.)	$2.0\text{--}4.4 \times 2.2\text{--}3.6\text{ }\mu\text{m}$ on PDA	$4.3 \times 2.8\text{ }\mu\text{m}$ on PDA

N/A, not available in the literature; PDA: Potato dextrose agar; MEA: Malt extract agar; SNA: Synthetic nutrient poor agar; CMAD: Corn meal agar with 1% dextrose.
^a Fungal strain used in this study, ^b Source of description [16].

Molecular phylogeny of KNU-4-KH1

After the sequence analysis of KNU-4-KH1, 549 bp from the ITS region, 586 bp from *TEF1α*, and 459 bp from *CAL* were obtained. BLAST analysis of the ITS sequences indicated a maximum identity of 100% with the strains of *Trichoderma ivoriense* (CBS 125734^T, BJ598-1). *TEF1α* showed maximum 98.7% and 99.3% similarities with the CBS 125734^T and BJ598-1 strains of *T. ivoriense*, respectively. In addition, the *CAL* gene showed the maximum 99.6% similarity to *T. ivoriense* CBS 125734^T. Phylogenetic analysis of the combined dataset of ITS, *TEF1α*, and *CAL* revealed that the strain from this study and the reference strain of *T. ivoriense* CBS 125734^T clustered together with a high bootstrap value (100%) in the NJ phylogenetic tree (Fig. 6). Thus, KNU-4-KH1 was identified as *T. ivoriense*.

The genus *Trichoderma* is found in various parts of the world on various substrates. The physicochemical mechanisms used by *Trichoderma* spp. to obtain nutrients allow them to thrive in a wide variety of substrates and environmental settings [12]. There are multiple species in the genus *Trichoderma*, making it hyperdiverse and with a global reach, spreading over many different substrates and locations [32]. This genus has proven to be useful in agriculture (biocontrol of plant pathogens), industry (prevention of pollution), and environmental protection [33,34]. Many researchers worldwide are interested in this group owing to its diversity and the resources it continues to explore. However, some species are responsible for the spread of green mold illness during mushroom farming [35] and even serve as opportunistic human diseases [36]. Although the genus *Trichoderma* serves as one of the best bioagents effective against a wide range of pathogens, the exact role of *T. ivoriense* as a biological control agent remains unknown. Therefore, the biological control activity of the isolated strain *T. ivoriense* requires further examination. To the best of our knowledge, this is the first report of *T. ivoriense* in Korea.

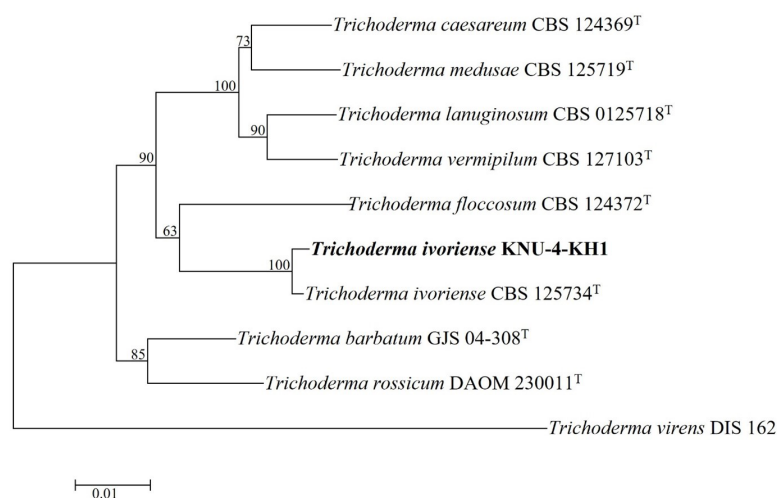


Fig. 6. Neighbor-joining phylogenetic tree of the strain KNU-4-KH1 based on the combined sequences (ITS+*TEF1α*+*CAL*), showing the relationships between *Trichoderma ivoriense* with the closest species of *Trichoderma*. *Trichoderma virens* DIS 162 was used as an outgroup. Numbers above the branches represent the bootstrap values obtained for 1,000 replicates. The isolated strain of this study is indicated in bold. Bar, 0.01 substitutions per nucleotide position. ITS, internal transcribed spacer of nuclear ribosomal DNA; *TEF1α*, translation elongation factor 1- α gene; *CAL*, calmodulin gene.

CONFLICT OF INTERESTS

The authors declare no conflict of interests.

ACKNOWLEDGMENTS

This study was supported by grants from the National Institute of Biological Resources (NIBR), funded by the Ministry of Environment of the Republic of Korea (NIBR201801105; NIBR201902112).

REFERENCES

1. Eriksson OE, Winka K. Supraordinal taxa of Ascomycota. *Myconet* 1997;1:1-16.
2. Maharachchikumbura SSN, Hyde KD, Jones EB, McKenzie EHC, Bhat JD, Dayarathne MC, Huang SK, Norphanphoun C, Senanayake IC, Perera RH, et al. Families of Sordariomycetes. *Fungal Divers* 2016;79:1-317.
3. Udayanga D, Castlebury LA, Rossman AY, Chukeatirote E, Hyde KD. Insights into the genus *Diaporthe*, phylogenetic species delimitation in the *D. eres* species complex. *Fungal Divers* 2014;67:203-29.
4. Mostert L, Crous PW, Kang JC, Phillips AL. Species of *Phomopsis* and a *Libertella* sp. occurring on grapevines with specific reference to South Africa: morphological, cultural, molecular and pathological characterization. *Mycologia* 2001;93:146-67.
5. Santos JM, Correia VG, Phillips AJL. Primers for mating-type diagnosis in *Diaporthe* and *Phomopsis*: their use in teleomorph induction in vitro and biological species definition. *Fungal Biol* 2010;114:255-70.
6. Gomes RR, Glienke C, Videira SIR, Lombard L, Groenewald JZ, Crous PW. *Diaporthe*: a genus of endophytic, saprobic and plant pathogenic fungi. *Persoonia* 2013;31:1-41.
7. Locquin-Linard M. *Faurelina*, nouveau genre d'Ascomycètes (Chadefaudiellaceae?). *Rev Mycol* 1975;39:125-9.
8. Parguey-Leduc A, Locquin-Linard M. L'ontogénie et la structure des périthèces de *Faurelina fimigenes* Locquin-Linard. *Rev Mycol* 1976;40:161-75.
9. von Arx JA. Notes on *Microascaceae* with the description of two species. *Persoonia* 1978;10:23-31.
10. Tang AMC, Jeewon R, Hyde KD. Phylogenetic utility of protein (RPB2, beta-tubulin) and ribosomal (LSU, SSU) gene sequences in the systematics of Sordariomycetes (Ascomycota, Fungi). *Antonie van Leeuwenhoek* 2007;91:327-49.
11. Qin WT, Zhuang WY. Seven wood-inhabiting new species of the genus *Trichoderma* (Fungi, Ascomycota) in Viride clade. *Sci Rep* 2016;6:27074.
12. Jaklitsch WM. European species of *Hypocrea* Part I. The green-spored species. *Stud Mycol* 2009;63:1-91.
13. Druzhinina IS, Kubicek CP. Species concepts and biodiversity in *Trichoderma* and *Hypocrea*: from aggregate species to species clusters? *J Zhejiang Univ Sci* 2005;B6:100-12.
14. von Arx JA, Mukerji KG, Singh N. *Faurelina indica* sp.nov. *Sydowia* 1981;34:39-41.
15. Nirenberg H. Untersuchungen u"ber die morphologische und biologische differenzierung in der *Fusarium-sektion* Liseola. *Mitt Biol Bundesanst Land-u Forstwirtschaft Berlin-Dahlem* 1976;169:1-17.

16. Zhang GZ, Zhang XJ, Zhou FY, Wang JN, Wu XQ, Zhao XY, Xie XY, Zhou HZ. Two Chinese new records of the genus *Trichoderma* in the *Stromaticum* clade. *Microbiology China* 2018;45:1120-6.
17. Gardes M, Bruns TD. ITS primers with enhanced specificity for Basidiomycetes—application to the identification of mycorrhizae and rusts. *Mol Ecol* 1993;2:113-8.
18. White T, Bruns T, Lee S, Taylor JW. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ, editors. *PCR protocols: a guide to methods and applications*. New York: Academic Press, Inc.; 1990. p. 315-22.
19. Vilgalys R, Hester M. Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *J Bacteriol* 1990;172:4238-46.
20. Carbone I, Kohn LM. A method for designing primer sets for speciation studies in filamentous Ascomycetes. *Mycologia* 1999;91:553-6.
21. Jaklitsch WM, Komon M, Kubicek CP, Druzhinina IS. *Hypocrea voglmayrii* sp. nov. from the Austrian Alps represents a new phylogenetic clade in *Hypocrea/Trichoderma*. *Mycologia* 2005;97:1365-78.
22. Glass NL, Donaldson G. Development of primer sets designed for use with PCR to amplify conserved genes from filamentous ascomycetes. *Appl Environ Microbiol* 1995;61:1323-30.
23. O'Donnell K, Cigelnik E. Two divergent intragenomic rDNA ITS2 types within a monophyletic lineage of the fungus *Fusarium* are nonorthologous. *Mol Phylogenet Evol* 1997;7:103-16.
24. Kimura M. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *J Mol Evol* 1980;16:111-20.
25. Kumar S, Stecher G, Tamura K. MEGA7: molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Mol Biol Evol* 2016;33:1870-4.
26. Udayanga D, Liu XZ, McKenzie EHC, Chukeatirote E, Bahkali AHA, Hyde KD. The genus *Phomopsis*: biology, applications, species concepts and names of common phytopathogens. *Fungal Divers* 2011;50:189-225.
27. Huang F, Hou X, Dewdney MM, Fu Y, Chen G, Hyde KD, Li H. *Diaporthe* species occurring on citrus in China. *Fungal Divers* 2013;61:237-50.
28. Sebastianes FLS, Lacava PT, Fávoro LCL, Rodrigues MBC, Araújo WL, Azevedo JL, Pizzirani-Kleiner AA. Genetic transformation of *Diaporthe phaseolorum*, an endophytic fungus found in mangrove forests, mediated by *Agrobacterium tumefaciens*. *Curr Genet* 2012;58:21-33.
29. Iriart I, Binois R, Fior R, Blanchet D, Berry A, Cassaing S, Amazan E, Papot E, Carme B, Aznar C, et al. Eumycetoma caused by *Diaporthe phaseolorum* (*Phomopsis phaseoli*): a case report and a mini-review of *Diaporthe/Phomopsis* spp. invasive infections in humans. *Clin Microbiol Infect* 2011;17:1492-4.
30. Santos PJ, Savi DC, Gomes RR, Goulin EH, Da Costa Senkiv C, Tanaka FA, Almeida ÁM, Galli-Terasawa L, Kava V, Glienke C. *Diaporthe endophytica* and *D. terebinthifolii* from medicinal plants for biological control of *Phyllosticta citricarpa*. *Microbiol Res* 2016;186:153-60.
31. Melo RFR, Miller AN, Maia LC. First records of *Faurelina* in the Neotropics. *Mycotaxon* 2016;131:679-85.
32. Chaverri P, Samuels GJ. *Hypocrea/Trichoderma* (Ascomycota, Hypocreales, Hypocreaceae): Species with green ascospores. *Stud Mycol* 2003;48:1-116.

33. Saravanakumar K, Kathiresan K. Bioconversion of lignocellulosic waste to bioethanol by *Trichoderma* and yeast fermentation. 3 Biotech 2014;4:493-9.
34. Adnan LA, Sathishkumar P, Yusoff AR, Hadibarata T, Ameen F. Rapid bioremediation of Alizarin Red Sand Quinizarine Green SS dyes using *Trichoderma lixii* F21 mediated by biosorption and enzymatic processes. Bioprocess Biosyst Eng 2017;40:85-97.
35. Komon-Zelazowska M, Bissett J, Zafari D, Hatvani L, Manczinger L, Woo S, Lorito M, Kredics L, Kubicek CP, Druzhinina IS. Genetically closely related but phenotypically divergent *Trichoderma* species cause green mold disease in oyster mushroom farms worldwide. Appl Environ Microbiol 2007;73:7415-26.
36. Sandoval-Denis M, Sutton DA, Cano-Lira J, Genéa J, Fothergill AW, Wiederhold NP, Guarro J. Phylogeny of the clinically relevant species of the emerging fungus *Trichoderma* and their antifungal susceptibilities. J Clin Microbiol 2014;52:2112-25.