RESEARCH NOTE

Stem Rot of Gondre Caused by *Rhizoctonia solani* AG-2-2(IV)

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ABSTRACT

Stem rot symptoms were observed in Gondre (*Cirsium setidens*) plants growing in a vinyl greenhouse in Taebaek, Korea during a disease survey in June 2022. The plants presented with dark brown to black rot on the stems at or above the soil line. Severely diseased plants displayed wilt and blight. Disease incidence among these plants ranged from 1 to 5%. Three isolates of *Rhizoctonia* sp. were obtained from the stem lesions of diseased plants. All isolates were identified as *Rhizoctonia solani* AG-2-2(IV) based on the morphological and cultural characteristics, results of the anastomosis test, and phylogenetic analysis. The pathogenicity of the isolates to Gondre plants was confirmed using an artificial inoculation test. The lesions induced by the inoculation test were similar to those observed in the investigated vinyl greenhouse. Here, we report a case of *R. solani* AG-2-2(IV) causing stem rot in Gondre.

Keywords: Anastomosis, Gondre, Cirsium setidens, Rhizoctonia solani AG-2-2(IV), Stem rot

Gondre [*Cirsium setidens* (Dunn) Nakai], also known as Korean thistle, belongs to the family Asteraceae and is native to Korea [1]. The plant is a perennial plant that grows naturally in montane and submontane areas, that is cultivated as a wild vegetable by farmers in Korea, and its young leaves can be cooked and seasoned. In June 2022, we surveyed the disease occurrence in wild vegetables cultivated in Taebaek, Korea. During the disease survey, stem rot symptoms were observed in Gondre plants growing in a vinyl greenhouse. Symptoms formed on the stems at or above the soil line, and the rotted lesions were dark brown to black (Fig. 1A and B). Severely diseased plants were wilted and blighted. We investigated stem rot outbreaks at three sites in the vinyl greenhouse, and after studying 100 plants per site, the disease incidence among the plants ranged from 1 to 5%.

Fungal isolates were obtained from the diseased stem lesions of Gondre plants using a previously described method [2]. Three isolates (RS-2207, RS-2209, and RS-2213) of *Rhizoctonia* sp. from the analyzed stem lesions were examined for morphological characteristics using a compound microscope (Nikon Eclipse Ci-L, Japan). Hyphae branched near the distal septa of the hyphal cells and were constricted at the points of hyphal branches. Hyphal septa were formed at a short distance from the points of the hyphal branches. These isolates did not produce conidia and clamp connections on the hyphae. The isolates were identified as *Rhizoctonia solani* Kühn, as per previous morphological descriptions [3,4].

R. solani isolates were tested for the identification of anastomosis groups using tester isolates of R. solani



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Fig. 1. Stem rot symptoms of Gondre plants. (A and B) Symptoms observed in the vinyl greenhouse. (C) Symptoms induced by artificial inoculation test with *Rhizoctonia solani* AG-2-2(IV) RS-2209. The white arrow indicates the lesion formed on the stem. (D) A non-inoculated plant (control).

(AG-1 through AG-5) as previously described [5,6]. All the tested isolates were classified as *R. solani* AG-2-2. The anastomosis reactions between the tested and tester isolates are shown in Fig. 2A. Colonies of the isolates cultured on potato dextrose agar (PDA) were light to dark brown in color (Fig. 2B). Sclerotia were absent or rarely formed on the medium. The isolates did not grow on PDA at 35°C. The cultural characteristics of the isolates were similar to those of the *R. solani* type IV cultures, which were described in a previous study [7].

To validate the identification results obtained via the morphological and cultural characteristics, and the results of the anastomosis tests, a phylogenetic analysis was conducted. The DNA templates of the three isolates were extracted using a previously described method [8], with slight modifications. Internal transcribed spacer regions 1 & 2 and intervening 5.8S nrDNA (ITS) were investigated using primers ITS1 and ITS4 [9]. Polymerase chain reaction (PCR) amplification was performed using the DNA Free-Multiplex Master Mix (Cellsafe, Korea) following the manufacturer's instructions. The amplifying condition included an initial denaturation at 95 °C for 5 min, followed by 35 cycles of denaturation at 95 °C for 30 sec, annealing at 55 °C for 60 sec, extension at 72 °C for 30 sec, and a final extension at 72 °C for 10 min. The PCR products were purified using a Universal DNA Purification Kit (Tiangen, China) following the manufacturer's instructions. Purified PCR products were sequenced at Bionics (Seoul, Korea). The obtained sequences were aligned using MUSCLE [10] and modified, when necessary, using MEGA 7 software [11].



Fig. 2. Anastomosis test and cultural appearance of *Rhizoctonia solani* RS-2209 from Gondre plants. (A) Anastomosis reactions between the tested isolate (td) and the tester isolate (tr) of *R. solani* AG-2-2(IV) observed using a compound microscope. The arrows indicate points of hyphal anastomosis. (B) A colony of the isolate grown on potato dextrose agar at 25° C for 10 days.

A phylogenetic tree was constructed by the neighbor-joining method with the maximum composite likelihood model, and 1,000 bootstrap replicates were performed. *Ceratobasidium* sp. (MAFF 237401) was used as an outgroup taxon. Sequence data of reference strains were obtained from the previous studies and NARO Genebank [12-24]. Phylogenetic analysis revealed that all the isolates were *R. solani* AG-2-2(IV) (Fig. 3). The nucleotide sequences of the three isolates were deposited in the NCBI GenBank under the accession numbers OQ991158–OQ991160.

Three isolates of *R. solani* AG-2-2(IV) were tested for pathogenicity to Gondre plants using an artificial inoculation test. Mycelial disks (10 mm in diameter) cut from the margins of actively growing cultures of each isolate on PDA were placed on the stems at the soil surface level of 5-month-old Gondre plants grown in circular plastic pots (height: 13.5 cm; upper diameter: 15 cm; lower diameter: 10 cm) in a vinyl greenhouse. Inoculated plant pots were placed in plastic boxes (72 cm \times 54 cm \times 45 cm) under 100% relative humidity at room temperature (24–28°C) for 7 days. Thereafter, the inoculated plant pots were taken out of the plastic boxes and kept under alternating cycles of 12 hr LED (light emitting diode) light and 12 hr darkness in a cultivation room at 24–28°C for 5 days. The inoculation test was carried out three times. The results of pathogenicity tests were checked for induced stem rot symptoms 12 days after inoculation. All the tested isolates induced stem rot symptoms in the inoculated plants (Fig. 1C), but no symptoms were observed in the control plants (Fig. 1D). The lesions induced by the inoculation test were similar to those observed in the vinyl greenhouse investigated during the survey. Furthermore, inoculated isolates were successfully detected in these lesions.

Several diseases, including powdery mildew and rust, have been reported in Gondre [25]. However, no prior reports of diseases caused by *R. solani* on this plant were noted. *R. solani* AG-2-2(IV) is known to cause stem rot in *Angelica jaluana* Nakai and *Gypsophila elegans* M. Bieb. in Korea [26,27]. Here, we report a case of *R. solani* AG-2-2(IV) causing stem rot in Gondre.



Fig. 3. Phylogenetic tree based on the sequences of internal transcribed spacer regions 1 & 2 and intervening 5.8S nrDNA of the three isolates (RS-2207, RS-2209, and RS-2213) from Gondre plants and reference strains. The reference sequence data were obtained from the NCBI GenBank database. The tree was generated using the neighbor-joining method with the maximum composite likelihood model in MEGA version 7 software. The bootstrap support values \geq 50 are shown at the nodes. The scale bar represents the number of nucleotide substitutions per site.

CONFLICT OF INTERESTS

No conflict of interest was reported by the authors.

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