

## RESEARCH NOTE

# Leaf Blight Caused by *Curvularia intermedia* on the Invasive Weed *Lactuca serriola* in Korea

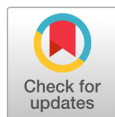
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## ABSTRACT

Prickly lettuce (*Lactuca serriola*), an invasive annual plant, poses a significant threat to the agricultural systems of many countries, including Korea. In 2020, leaf blight symptoms were observed in the prickly lettuce populations of various farms across Korea. Detailed morphological and molecular sequence analyses revealed that the disease was caused by the fungus, *Curvularia intermedia*. A pathogenicity test confirmed that the fungus can cause the same symptoms in healthy prickly lettuce, thereby fulfilling Koch's postulates. To the best of our knowledge, this is the first report of *C. intermedia* causing leaf blight on *L. serriola* in Korea, suggesting its potential as a biocontrol agent for this weed. However, further investigations are necessary to determine its ecological impact to prevent any non-target effects.

**Keywords:** Biological control, *gpd* (glyceraldehyde-3-phosphate dehydrogenase), *Lactuca scariola*, Prickly lettuce



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*Lactuca serriola* L. (syn. *L. scariola* L.), commonly known as prickly lettuce, is an annual plant belonging to the family Asteraceae. This plant is native to Europe and has become widely naturalized in temperate regions worldwide. It poses a severe risk to agricultural systems, especially in reduced- or no-tillage cultivation methods, which are commonly used for cereals, pulses, soybeans, and wheat [1,2]. *L. serriola* causes a drastic decline in crop yield by 60-80% at high population density and significantly affects the quality of grains during harvest [1,3,4].

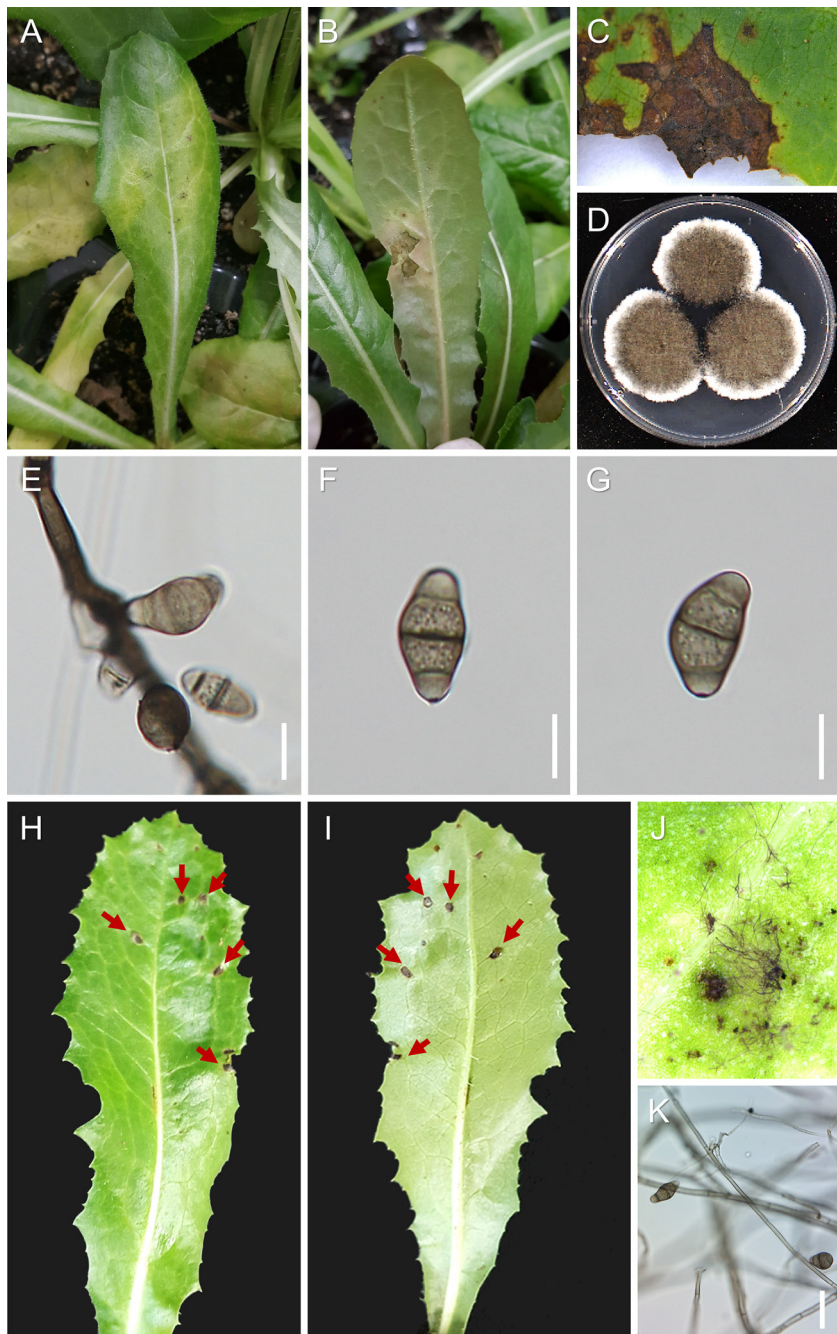
Prickly lettuce was accidentally introduced to Korea in the late 1970s [5,6] and has since rapidly spread to various agricultural and natural habitats across the country [7]. Owing to its adverse effects, the Korean Ministry of Environment has designated this weed as a harmful plant that disturbs the balance of the ecosystem [6]. Prickly lettuce seeds are highly adaptable and able to germinate at temperatures as low as 10°C albeit at a slower speed [3], contributing to their rapid spread across Korea. Furthermore, the survival of this weed in harsh and barren environments, such as the roadsides, in Korea is facilitated by endophytic bacteria, which increase its drought resistance capacity [8].

In June 2020, we observed leaf blight symptoms on prickly lettuce populations invading several farms in Boryeong-si (36°22'46" N, 126°35'37" E) in Korea, with a disease incidence of 30–40% among the inspected plants. Disease symptoms began as small light yellow-to-brown adaxial leaf spots, which gradually coalesced into larger irregular lesions and eventually withered (Fig. 1A and B). Twenty symptomatic leaves were collected and used for fungal isolation. Sections (5 × 5 mm) of these leaves were surface-sterilized (70% ethanol for 1 min, 0.05% NaOCl for 30 s, and 70% ethanol for 1 min), rinsed twice with sterile water, and placed on a potato dextrose agar (PDA) plate with streptomycin (50 µg/mL). The plant sections were incubated in the dark at 25°C until fungal growth was observed. After three days, 15 morphologically similar isolates were selected, and a pure culture was obtained by transferring single hyphal tips to a new PDA plate. Colonies on PDA appeared gray-to-dark brown and velvety, with approximately 4 cm in diameter after three days (Fig. 1C). A sample culture was deposited in the Korean Agricultural Culture Collection (KACC 49814).

The causative agent was identified through a combination of morphological and molecular sequence analyses. Detailed microscopic investigations were performed on the conidiophores and conidia formed on KACC 49814 PDA. Samples were examined and imaged using the Zeiss Imager M2 AX10 microscope (Carl Zeiss, Jena, Germany) equipped with an AxioCam 512 camera (Carl Zeiss). Conidiophores were brown-to-pale brown, simple but rarely branched, septate, thick-walled, and measured (53–) 80 to 128 (–170) × (3–) 4 to 7 (–8) µm (n=30). Conidia (Fig. 1D–F) were dark brown, obclavate to ellipsoidal or fusiform, mostly 3-septate, straight or slightly curved, with a rough verrucose surface, and measured (23–) 28 to 33.3 (–37) × (10–) 12 to 15 (–17) µm (n=50). Terminal cells were smaller, paler, and less verrucose than the central cells. Their morphological characteristics matched those of *Curvularia intermedia* Boedijn [9].

To confirm the morphological identification, molecular sequence analysis was performed. Genomic DNA was extracted from the fungal conidia and hyphae harvested from KACC 49814 using the DNeasy Plant Mini Kit (Qiagen, Germantown, MD, USA). Then, the internal transcribed spacer (ITS) and glyceraldehyde-3-phosphate dehydrogenase (*gpd*) [10] were PCR-amplified and sequenced using a DNA sequencing service (Macrogen, Seoul, Korea) with the primers used for amplification. The resulting sequences were edited using the DNASTAR Lasergene software package (DNASTAR, Madison, WI, USA) and deposited in GenBank (accession nos. MZ373300 for ITS and MZ435744 for *gpd*). A BLASTn search revealed that the Korean isolate had high sequence similarities of 100% and 99.6% with *C. intermedia* CBS 334.64 for ITS (HG778991) and *gpd* (LT715828), respectively.

Pathogenicity assays were performed twice by spraying a conidial suspension ( $1.9 \times 10^5$  conidia/mL) onto each leaf of 20 healthy *Lactuca serriola* (four-week-old) plants. Twenty control plants were sprayed with sterile water. All plants were kept in a growth chamber at 23°C and 80% relative humidity under a 12-h day/night cycle. One week after inoculation, initial symptoms began to emerge as small foliar spots, which turned yellow to brown in two weeks, leading to leaf dieback and necrosis (Fig. 1G–I). Control plants



**Fig. 1.** Leaf blight on prickly lettuce (*Lactuca serriola*) caused by *Curvularia intermedia*. (A and B) Symptoms observed on the upper (A) and lower (B) surfaces of an infected leaf. (C) A close-up of a leaf blight lesion on the upper leaf surface. (D) Colony morphology observed on the potato dextrose agar (PDA) plate after incubation for a week in darkness at 25°C. (E-G) Conidia. (H and I) Pathogenicity assays showing typical brown spots (arrows) on the front (H) and backside (I) of an inoculated leaf. (J) Enlarged region of the inoculated leaf showing symptoms. (K) Conidia re-isolated from an inoculated leaf. Scale bars: 10 µm (E-G) and 30 µm (K).

were symptom-free. The same fungus was successfully reisolated from the artificially inoculated plants (Fig. 1J), thereby fulfilling Koch's postulates.

In the last decade, several fungal pathogens, including *Alternaria alternata* [11], *Golovinomyces orontii* (recently reclassified under *G. bolayi*) [12], *Fusarium fujikuroi* [13], and *Septoria lactucae* [14], have been reported on *L. serriola* in Korea. Additionally, oomycetes, such as *Bremia lactucae* [15], have also been documented. To the best of our knowledge, this is the first report of leaf blight caused by *C. intermedia* on *L. serriola* in Korea. *C. intermedia* affects the economically important crops and grasses as well as weeds of lawns and turfgrasses [16]. Furthermore, it causes leaf spots on *Zea mays* in Korea [17].

Results of this study suggest *C. intermedia* as an effective biocontrol agent for *L. serriola*. This pathogen was previously proposed as a potential microbial herbicide for large crabgrass (*Digitaria sanguinalis*) [18]. In addition, use of *C. intermedia* as a microbial herbicide against crabgrass and other susceptible species has been detailed in two U.S. patents [19,20]. However, its potential non-target effects require further investigation to prevent any adverse ecological impact.

## CONFLICT OF INTERESTS

The authors declared no conflicts of interest.

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## REFERENCES

1. Chadha A, Florentine S. Biology, ecology, distribution and control of the invasive weed, *Lactuca serriola* L. (wild lettuce): A global review. *Plants* 2021;10:2157.
2. Weaver S, Cluney K, Downs M, Page E. Prickly lettuce (*Lactuca serriola*) interference and seed production in soybeans and winter wheat. *Weed Sci* 2006;54:496-503.
3. Wu H, Asaduzzaman M, Shephard A, Hopwood M, Ma X. Germination and emergence characteristics of prickly lettuce (*Lactuca serriola* L.). *Crop Prot* 2020;136:105222.
4. Amor RL. Incidence and growth of prickly lettuce (*Lactuca serriola* L.) in dryland crops in the Victorian Wimmera. *Plant Prot Q* 1986;1:148-51.
5. Yim YJ, Jeon ES. Distribution of naturalized plants in the Korean peninsula. *Kor J Bot* 1980;23:69-83.
6. Kim YH, Kil JH, Hwang SM, Lee CW. Spreading and distribution of *Lactuca scariola*, invasive alien plant, by habitat types in Korea. *Weed Turfgrass Sci* 2013;2:138-51.
7. Lee IY, Kim SH, Lee YH, Hong SH. Occurrence characteristics and management plans of *Lactuca scariola* L., an ecosystem disturbance plant. *Korean J Environ Biol* 2022;40:239-46.

8. Jeong S, Kim TM, Choi B, Kim Y, Kim E. Invasive *Lactuca serriola* seeds contain endophytic bacteria that contribute to drought tolerance. *Sci Rep* 2021;11:13307.
9. Sivanesan A. Graminicolous species of *Bipolaris*, *Curvularia*, *Drechslera*, *Exserohilum* and their teleomorphs. *Myc Papers* 1987;158:1-261.
10. Berbee ML, Pirseyedi M, Hubbard S. *Cochliobolus* phylogenetics and the origin of known, highly virulent pathogens, inferred from ITS and glyceraldehyde-3-phosphate dehydrogenase gene sequences. *Mycologia* 1999;91:964-77.
11. Kim BR, Choi YJ. *Alternaria alternata* causing leaf spot on *Lactuca serriola* in Korea. *Plant Dis* 2020;104:2293.
12. Park MJ, Hong SH, Cho SE, Park JH, Shin HD. First report of powdery mildew caused by *Golovinomyces orontii* on invasive weed *Lactuca serriola* in Korea. *Plant Dis* 2015;99:889.
13. Kim BR, Choi YJ. *Fusarium fujikuroi* causing fusarium wilt of *Lactuca serriola* in Korea. *Plant Dis* 2021;105:502.
14. Kim BR, Choi YJ, Shin HD. First report of leaf spot caused by *Septoria lactucae* on *Lactuca serriola* in Korea. *Plant Dis* 2020;104:581.
15. Lee JA, Kim B, Lee DJ, Choi YJ. *Bremia lactucae* causing downy mildew on *Lactuca serriola* in Korea. *Kor J Mycol* 2021;49:379-83.
16. Farr DF, Rossman AY. Fungal databases [Internet]. Beltsville, MD: U.S. National Fungus Collections, ARS, USDA; 2023 [cited 2023 Aug 8]. Available from: <http://nt.ars-grin.gov/fungaldatabases/>.
17. KSPP (The Korean Society of Plant Pathology). List of plant diseases in Korea. 6 ed. Seoul: The Korean Society of Plant Pathology; 2022.
18. Michael Tilley A, Lynn Walker H. Evaluation of *Curvularia intermedia* (*Cochliobolus intermedium*) as a potential microbial herbicide for large crabgrass (*Digitaria sanguinalis*). *Biol Control* 2002;25:12-21.
19. Walker HL, Tilley AM. Control of crabgrass with a fungal pathogen. United States Patent No. 5,635,444. 1997.
20. Walker HL, Tilley AM. Control of crabgrass with a fungal pathogen. United States Patent No. 5,952,264. 1999.