RESEARCH ARTICLE

# **Morphological and Phylogenetic** Characteristics of Tuber himalayense **Collected from Rhizosphere of Quercus** dentata in Korea

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# ABSTRACT

We collected the ascomata of Tuber species from the rhizosphere of Quercus dentata in Danyang, Korea. We observed the morphological characteristics of ectomycorrhizal roots and ascomata, and identified the species based on the results of the phylogenetic analysis conducted using the DNA sequences of an internal transcribed spacer, a large-subunit rDNA, translation elongation factor  $1-\alpha$  DNA (TEF1), and MAT. Finally, we identified the fungal species as Tuber himalayense B.C. Zhang & Minter, which has not been recorded previously in Korea. We evaluated the morphological characteristics and conducted phylogenetic analysis of the ascoma and mycorrhiza (associated with Q. dentata) of T. himalayense.

Keywords: Ascoma, Ectomycorrhiza, Fruiting body, Truffle, Tuber himalayense

# INTRODUCTION

Truffles are a group of fungi that produce hypogeous ascocarps; they belong to the genus *Tuber* (Ascomycota, Pezizales) [1]. Tuber spp. have ectomycorrhizal (ECM) associations with woody plants such as Quercus, Pinus, and Corylus [2,3]. Truffles have been widely used as edible mushrooms in Europe for a long time owing to their unique fragrance and flavor [1].

Tuber himalayense B.C. Zhang & Minter was first reported in 1988 [4] and its morphological characteristics were largely consistent with those of T. indicum, known as Asian black truffle; furthermore, it shared morphological and phylogenetical similarities with European black truffle Tuber melanosporum. However, the fruiting bodies of T. himalayense are smaller than those of T. indicum and polygonal warts attached to the outer side of the peridium has been recorded to be regular compared to those of T. indicum [3,4]. As T. himalayense and T. indicum are systematically very close, their taxonomic distinction has not been clarified and the two species have been considered as one or as T. indicum complex [5]. Recently, T.



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*himalayense* and *T. indicum* have been distinguished at the species level using DNA sequences of the MAT genes, which determine the mating type of *Tuber* spp. [6,7].

The ascoma of only two *Tuber* spp. (*Tuber aestivum* subsp. *uncinatum* and *Tuber huidongense*) have been reported in Korea [8, 9]. Herein, we describe the morphological characteristics of the fruiting bodies of *T. himalayense* found in the rhizosphere of *Quercus dentata* Thunb. (an oak tree) in Korea and their mycorrhizal roots. We determined whether *T. himalayense* systematically differs from *T. indicum* using several molecular markers, including the MAT genes.

## MATERIALS AND METHODS

*The as*comata of *T. himalayense* were collected from the rhizosphere of *Q. dentata* in Danyang, Korea. The morphological characteristics of ascoma, asci, and ascospores were examined under a light microscope. For molecular identification, genomic DNA was extracted from the ascoma and ITS1/ITS4 primers [10] and LR0R/LR16 primers were used to amplify internal transcribed spacer (ITS) regions and large subunits (LSU) of ribosomal DNA (rDNA) [11], respectively. EF1 $\alpha$  Tuber-f and EF1 $\alpha$  Tuber-r, and Tuber-r [12] were also used to amplify the translation elongation factor 1- $\alpha$ . Additionally, the i3/il2 primers and i5/ il3 primers [7] were used to amplify MAT 1-1-1 and MAT 1-2-1 DNA for phylogenetic analysis with *T. indicum*. After PCR amplification, DNA sequences were analysed (SolGent Co., Ltd., Daejeon, Korea). Sequence similarities were determined using BLAST available from the National Center for Biological Information (NCBI). Phylogenetic trees were constructed using the neighbor-joining method with MEGA7 software [13].

The roots of *Q. dentata* were collected from the same sites in Danyang where the ascoma were collected. The morphological and anatomical characteristic of ECM were examined under a stereomicroscope and light microscopes. Genomic DNA was extracted from the ECM root tips using DiaStar<sup>TM</sup> Direct Lysis Buffer (SolGent Co., Ltd., Daejeon, Korea). The ITS region of the rDNA was amplified using the ITS1F/ITS4 primers and identified through phylogenetic analysis of the sequences.

### **RESULTS AND DISCUSSION**

#### Taxonomy

**Tuber himalayense** B.C. Zhang & Minter, Transactions of the British Mycological Society 91 (4): 595 (1988) [MB#134661] (Fig. 1; Table 1)

Globose to subglobose, black to blackish brown,  $20-35 \times 17-28$  mm in diameter (Figs. 1A and 1B), polygonal ornamentation on the surface (Fig. 1C),  $0.3-0.5 \times 0.2-0.3$  mm in diameter. Peridium divided into two distinct layers: an outer layer, dark brown to black, and (222.90-)267.51(-297.29) µm in thickness, and an inner layer, purplish brown to reddish brown, and (113.71-)135.01(-165.16) µm in thickness. Gleba initially greyish beige and dark brown in the part with mature ascospores (Fig. 1D). Asci hyaline, irregular,

with rounded margin, ellipsoidal to conical, differing with the number of ascospores (2-4) in each ascus, (48.33-) 61.07 (-76.28)  $\times$  (32.09-) 49.72 (-62.20)  $\mu$ m in diam. (Figs. 1E and 1F). Ascospore yellowish brown to dark brown, ellipsoid or fusiform, and (16.89-) 25.37 (-35.79)  $\times$  (12.78-) 18.00 (-24.24)  $\mu$ m in diameter (Figs. 1G and 1H). The reticular structure on the surface of the mature ascospore resembles a turtle shell, with several spines on the external surface. These spines narrowed away from the basal structure and were usually straight although a few were curved, and (2.68-) 4.47 (-6.75)  $\mu$ m in length.

**Specimen examined.** Maepo-eup, Danyang-gun, Chungcheongbuk-do, Korea, October 16, 2020, *Tuber himalayense* B.C. Zhang & Minter, isolated from the rhizosphere of *Quercus dentata*, strain CB20001. GenBank accession numbers, MW393547 (ITS rDNA from ascoma), MW386847 (LSU rDNA from ascoma), MW810351(TEF1 from ascoma) and MW403878 (ITS rDNA from ECM roots).



**Fig. 1.** Morphological characteristics of *Tuber himalayense*. Ascoma (A, B), surface ornament (C), peridium and gleba (D) asci (E, F), and ascospores (G, H) Scale bars: B=5,000 μm, C, D=2,000 μm, E=50 μm, F=20 μm, G, H=10 μm.

Table 1. Morphological characteristics of Tuber himalayense ascoma from Danyang, Korea.

Characteristics	T. himalayense CB20001	T. himalayense [3,4]
Shape	Globose to subglobose, black to blackish	Irregular but globose, lobed surface, black,
	brown, irregular polygonal ornament	polygonal wart
Size	(20-35)×(17-28) mm in diameter	20-25 mm in diam.
Asci	(2-4) spored, hyaline, irregular, with rounded margin, ellipsoidal to conical, (48.33-)61.07(-76.28)×(32.09-)49.72(-62.20) $\mu m$ in diam.	(1-)2-4(-5) spored, mostly globose, colourless, 55-70 μm in diam.
Ascospores	Yellowish brown to dark brown, ellipsoid or fusiform, (16.89-)25.37(35.79)×(12.78-)18.00(-24.24) $\mu m$ in diam.	Brown to dark reddish brown, ellipsoid to subglobose, 28-45(-50)×23-40(-45) µm in diam.
Spines	Straight but sometimes curved, (2.68-)4.47(-6.75) µm in high	Up to 6 µm in high

#### **Phylogenetic analysis**

The phylogenetic analysis showed that the ascoma of CB20001 shared similarity of 98.87% with *T. himalayense* AB553388.1 for the ITS regions, 98.68% with *T. himalayense* AB553517.1 for the LSU regions, and 99.88% with *T. himalayense* AB553537.1 for the TEF1 regions. Phylogenetic analysis showed that the sequences for CB20001 was included in the same clade as *T. himalayense* (Fig. 2). Additionally, the sequence analysis of the *MAT* genes showed 100% similarity with *T. himalayense* LC312350.1 for the MAT 1-1-1 region and 100% similarity with *T. himalayense* LC312317.1 for the MAT 1-2-1 region. The phylogenetic analysis confirmed that these sequences formed a distinct clade with the sequences of *T. indicum* (Fig. 3).

#### Ectomycorrhiza of T. himalayense x Q. dentata

The ECM root tips were straight, velvety and brown to reddish brown (rarely red) (Figs. 4A-C). The cross section of the ECM showed the fungal mantle and Hartig net (Figs. 4D and 4E). The fungal mantle layer had an interlocking irregular synenchyma (Fig. 4F). The BLAST results showed that the ITS sequence of the ECM root tip is closely related to that of *T. himalayense* AB553393.1 (99.66% similarity); thus, it was identified as *T. himalayense* (Fig. 5).



0.02

**Fig. 2.** Neighbor-joining phylogenetic tree of *T. himalyense* CB20001 ascoma based on concatenated alignment of internal transcribed spacer (ITS), large subunit (LSU) rDNA, and translation elongation factor  $1-\alpha$  (TEF1) DNA sequences. *Cenococcum geophilum* was used as an outgroup. Numbers on branches indicate bootstrap values (1,000 replicated). The sequence from the present study is shown in bold.



0.02

**Fig. 3.** Neighbor-joining phylogenetic tree of *T. himalayense* based on the alignment of MAT 1-1-1 (A) and MAT 1-2-1 (B) sequences. *Tuber melanosporum* was used as the outgroup in both (A) and (B). Numbers on branches indicate bootstrap values (1,000 replicate). The sequences from the present study are shown in bold.



**Fig. 4.** Morphological characteristics of ectomycorrhiza colonized by *Tuber himalayense* from root of *Quercus dentata*. Mycorrhizal root tips (A-C); Transverse section of root tips stained with lactophenol cotton blue (D), outer mantle layer (E, F) Scale bars: A-C=500 µm, D=100 µm, E=50 µm, F=20 µm.



**Fig. 5.** Neighbor-joining phylogenetic tree of *T. himalayense* based on the alignment of internal transcribed spacer (ITS) rDNA sequences obtained from ectomycorrhizal root tips (MW403878) and ascoma of CB20001 (MW393547) in this study. *Cenococcum geophilum* was used as an outgroup. Numbers on branches indicate bootstrap values (1,000 replicates). Sequences from the present study are shown in bold.

The phylogenetic position of *T. himalayense* has not been clear due to its highly similar morphological characteristic with *T. indicum*. Chen *et al.* [14] classified *T. himalayense* and *T. indicum* into a *T. indicum* complex based on the phylogenetic analysis of LSU rDNA and beta-tubulin (TUB) DNA, whereas Qiao *et al.* [15] suggested classifying *T. indicum* and *T. himalayense* into different groups based on the analysis using simple sequence marker (SSRs). Recently, phylogenetic analysis using ITS, TUB, TEF1, and mating genes, MAT1-1-1 and MAT1-2-1, showed that the two species can be divided into separate clades and a new species, *T. longispinosum* was positioned between the clades Kinoshita *et al.* [9]. The ascoma of CB20001 collected from the rhizosphere of the oak tree was identified in this study using the ITS, LSU, and TEF1 regions and MAT genes, and the phylogenetic analysis validated the identification of CB20001 as *T. himalayense*. Additionally, the morphological characteristics of mycorrhizas of *T. himalayense* associated with *Q. dentata* were confirmed. The host plants of *T. himalayense* include *Quercus acutissima* Carruth, *Quercus serrata* Thunb. ex. Murray, and *Pinus densiflora* Siebold & Zucc. [9], however, in this study, *Q. dentata* was confirmed as a host plant of *T. himalayense*.

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# REFERENCES

- 1. Trappe JM. The orders, families, and genera of hypogeous Ascomycotina (truffles and their relatives). Mycotaxon 1979;9:297-340.
- Benucci GMN, Csorbai AG, Falini LB, Bencivenga M, Di Massimo G, Donnini D. Mycorrhization of *Quercus robur* L., *Quercus cerris* L. and *Corylus avellana* L. seedlings with *Tuber macrosporum* Vittad. Mycorrhiza 2012;22:639-46.
- García-Montero LG, Díaz P, Di Massimo G, García-Abril A. A review of research on Chinese Tuber species. Mycol Prog 2010;9:315-35.
- Zhang B, Minter D. Tuber himalayense sp. nov. with notes on Himalayan truffles. Trans Brit Mycol Soc 1988;91:593-7.
- 5. Wang YJ, Tan ZM, Zhang DC, Murat C, Jeandroz S, Le Tacon F. Phylogenetic and populational study of the *Tuber indicum* complex. Mycol Res 2006;110:1034-45.
- Belfiori B, Riccioni C, Paolocci F, Rubini A. Mating type locus of Chinese black truffles reveals heterothallism and the presence of cryptic species within the *T. indicum* species complex. Plos One 2013;8:e82353.
- Kinoshita A, Nara K, Sasaki H, Feng B, Obase K, Yang ZL, Yamanaka T. Using mating-type loci to improve taxonomy of the *Tuber indicum* complex, and discovery of a new species, *T. longispinosum*. Plos One 2018;13:e0193745.

- Shin KS, Park JS, Yoshimi S. Note on *Tuber aestivum* subsp. *uncinatum* newly recorded in Korea. Kor J Mycol 1995;23:10-3.
- Park H, Gwon JH, Lee JC, Kim HS, Oh DS, Eom AH. Report on *Tuber huidongense*, a truffle species previously unrecorded in Korea. Kor J Mycol 2020;48:505-10.
- Gardes M, Bruns TD. ITS primers with enhanced specificity for basidiomycetes-application to the identification of mycorrhizae and rusts. Mol Ecol 1993;2:113-8.
- Moncalvo JM, Lutzoni FM, Rehner SA, Johnson J, Vilgalys R. Phylogenetic relationships of agaric fungi based on nuclear large subunit ribosomal DNA sequences. Syst Biol 2000;49:278-305.
- Bonito G, Smith ME, Nowak M, Healy RA, Guevara G, Cázares E, Kinoshita A, Nouhra ER, Domínguez LS, Tedersoo L, et al. Historical biogeography and diversification of truffles in the Tuberaceae and their newly identified Southern Hemisphere sister lineage. Plos One 2013;8:e52765.
- Kumar S, Stecher G, Tamura K. MEGA7: Molecular evolutionary genetics analysis version 7.0 for bigger datasets. Mol Biol Evol 2016;33:1870-4.
- Chen J, Guo SX, Liu PG. Species recognition and cryptic species in the *Tuber indicum* complex. Plos One 2011;6:e14625.
- 15. Qiao P, Tian W, Liu Pg, Yu FQ, Chen J, Deng XJ, Wan SP, Wang R, Wang Y, Guo H. Phylogeography and population genetic analyses reveal the speciation of the *Tuber indicum* complex. Fungal Genet Biol 2018;113:14-23.